

Chapter 4

Role of cytochrome P450
polymorphisms in the development
of pulmonary drug toxicity
A case-control study in the Netherlands

P Wijnen, M Drent, P Nelemans, P Kuijpers, G Koek, C Neef, G Haenen, O Bekers

Drug Saf. 2008;31:1125-1134

Abstract

Background

Drug-induced pulmonary toxicity is a serious and expanding problem with often unknown aetiology. Many drugs are metabolized by cytochrome P450 (CYP) enzymes.

To establish whether allelic variation in CYP polymorphic genes contributes to variability in drug response and unexpected toxicity.

Study design

A case-control study was conducted. The cases consisted of patients with drug-induced interstitial lung disease (DI-ILD; n=59). Two control groups were used: one group of healthy volunteers (n=173) and one group of patients with idiopathic pulmonary fibrosis (IPF; n=110).

Results

Of the patients with DI-ILD 91.5% (54/59) had at least one of the studied variant genes compared with 70.5% (122/173, p<0.001) of the healthy volunteers and 69.1% (76/110, p<0.001) of the IPF patients. The percentage of individuals with one or more variant CYP genes was higher in DI-ILD group. Odds ratios were significantly increased and ranged from 3.25 to 40.8, indicating a significant association between the development of DI-ILD and the presence of one or more variant CYP genes.

Conclusion

DI-ILD appeared to be associated with the presence of at least one variant CYP allele. This study supports the potential usefulness of personalized medicine by genotyping aiming to improve efficacy, tolerability and drug safety.

Background

One may view the lung as bathed in two dissimilar environments: inhaled air and circulating blood. Both can carry noxious substances that may inflict damage to the lung. Air can deliver noxious particles and blood is the main supplier of most drugs, independent of the route of administration, and adverse respiratory reactions may follow by the most unlikely routes. Drugs can induce specific respiratory reactions, or the lungs may be affected as part of a generalized response. The most common form of drug-induced respiratory disease is diffuse interstitial lung disease (ILD). The drugs involved not only comprise prescribed and over-the-counter-drugs, but also illicit drugs, herbs, alcohol and ingredients of the diet. The list of compounds involved has grown rapidly over the last decades.^{1,2} Although drugs have been unequivocally identified as the cause in only a limited number of cases, it is important to acknowledge the potential role of medication in the development of ILD. This is due to the severity of the potentially irreversible damage to the lungs and the improvement that is often easily brought about by stopping administration of the drug involved.

The diagnosis of drug-induced ILD (DI-ILD) primarily rests on the link between drug intake and the subsequent development of respiratory symptoms. In some cases, the evidence that reactions in the lung are drug-induced is circumstantial. A straightforward interpretation is hampered when the damage is irreversible or when the symptoms are aggravated after stopping drug administration. Rechallenging the patient with the drug involved is frequently unethical and dangerous. Sensitive and specific tests are lacking. The key to diagnosis is a high clinical acuity.² All medication needs to be recorded and reviewed meticulously.

The molecular mechanism involved in the development of DI-ILD is still enigmatic. Inflammation initiated by drugs, drug metabolites or drug-induced free radical generation processes has been implicated.³ There is also emerging evidence for a role of genetic factors in the development of ILD such as differences in gene expression profiles between pulmonary fibrosis (IPF) and hypersensitivity pneumonitis.4,5

Cytochrome P450 (CYP) single nucleotide polymorphisms (SNPs) are one of the key factors known to cause a variation in drug response between individuals.³, ⁶⁻⁹ CYP is a super family of microsomal enzymes that metabolize various endogenous compounds and xenobiotics, including most drugs. Through CYP metabolism the chemical structure of drugs is changed, in general facilitating elimination of the drug from the body.^{3,7}

The risk for development of DI-ILD and clinical patterns vary depending on a variety of host and drug factors.² Several different phase I xenobioticmetabolizing CYP and phase II enzymes (i.e. conjugation enzymes including several transferases) are present in the human lung, possibly contributing to in situ activation. 10-14 Metabolism also affects the biological activity of the drug. The biological activity of the parent drug is usually superior to that of the metabolite, but there are several exceptions. Sometimes CYP metabolism yields very toxic metabolites, for example, with paracetamol (acetaminophen), benzo[a]pyrene and carbamazepine. 14 Occasionally, drugs may cause the formation of reactive oxygen species by uncoupling of the electron transport of the CYP system.¹⁵ These metabolites or reactive oxygen species may damage vital cellular components, such as proteins, lipids or DNA. 16 This might be the cause of clinically relevant drug-induced pulmonary toxicity. 12,17,18

Recently, we reported two cases of patients with interstitial pneumonia who developed cardiac failure following treatment with venlafaxine. ¹⁹ In both cases, a strong relationship between the development of the patients' illness and the initiation of venlafaxine treatment was identified. Furthermore, members of the CYP family are involved in the metabolism of venlafaxine. Therefore, we hypothesized that genetically or environmentally induced inter-individual differences in the expression of pulmonary biotransformation enzymes such as CYP may form the basis for, or contribute to, the risk of DI-ILD. 2,10,13,14

The aim of this study was to establish whether variation in CYP genes contributes to variability in individual drug response and toxicity. Therefore, the presence of the most clinically relevant variants of CYP genes (CYP2D6, CYP2C9 and CYP2C19 variants) was compared in a population of patients with DI-ILD with the presence of these variant genes in a population of patients with IPF and one of healthy volunteers.

Materials and methods

Setting and study population

The study was conducted as a case-control study. Between 2002 and 2006, 575 bronchoalveolar lavages (BAL) were performed on patients referred to the department of Respiratory Medicine of the Maastricht University Medical Centre (MUMC), Maastricht, The Netherlands, suspected of a non-infectious or infectious disorder. Out of these 575 cases, 51.1% appeared to have an infectious disorder, 17.8% had diffuse alveolar haemorrhage and 31.1% an ILD. Of this latter group, 59 (10.2%) met the criteria of DI-ILD. All of the 59 patients were Caucasian, used multiple drugs for various indications and did not have a history of any pulmonary disorder. The clinical presentation of druginduced pulmonary toxicity varied. Besides pulmonary symptoms, some patients (n=7; 12%) showed signs of other toxicity such as skin, cardiac and gastrointestinal involvement. The diagnosis DI-ILD was established by clinical presentation including dyspnoea and hypoxia, diffuse interstitial features on chest X-ray and a high-resolution CT scan as well as a BAL fluid (BALF) profile compatible with DI-ILD, excluding an infectious cause. ^{2,20,21} A lung biopsy was performed in 20% (n=12) of the patients. Clinical records were reviewed carefully and present and past drug use was documented. After reviewing the drug use of all DI-ILD patients, the most important groups appeared to be antihypertensive medication, β-adrenergic receptor antagonists, anti-arrythmic agents, antidepressants and anticoagulants. Every DI-ILD patient used a combination of at least two or more of these drugs. Withdrawal of the suspected causative drug(s) led to a favorable outcome in 75% of the patients. including an improvement of the respiratory symptoms, lung function test results, especially the diffusing capacity, and the chest X-ray abnormalities. In 25% of cases the damage was irreversible. Subjects were genotyped retrospectively for CYP polymorphisms and the drug(s) which are metabolized by the CYP enzymes that show genetic polymorphisms were identified.

The first control group consisted of 173 healthy Caucasian volunteers, recruited for method validation, who did not use medication nor had any relevant medical history, especially no history of pulmonary complaints.²² All healthy volunteers were hospital employees. This healthy volunteer control group was also used to establish the distribution of allele variants in the general population.

The second control group consisted of 110 Caucasian patients with IPF, known at our out-patient clinic and collected during the timeframe of this study, who also used medication for various indications. This group was selected as a control group, for it was expected that drug use did not play a substantial role in the pathogenesis of this disease. The diagnosis IPF was based on consistent clinical features, radiographic findings and BALF analysis results. According to the international consensus a biopsy was obtained from 50% of the IPF group, which confirmed the diagnosis histologically as being usual interstitial pneumonia.²³ Clinical records were reviewed carefully and present and past drug use was documented. All IPF patients used one or more drug(s). For the DI-ILD group as well as for the IPF group it was checked whether the drugs used were metabolized by polymorphic CYP enzymes.

The study was performed in accordance with the Declaration of Helsinki and its amendments. The protocol was approved by the local Medical Ethics Board of the MUMC. Written informed consent for participation in this study was obtained from all subjects.

Genotyping

DNA was obtained from all subjects by using venous EDTA anti-coagulated blood and isolating with a High Pure PCR Template Preparation Kit (Roche Mannheim, Germany) according Diagnostics, to the manufacturer's instructions.

In this study, genotyping was carried out for the following CYP allelic variants: CYP2C9*2 C430T, CYP2C9*3 A1075C, CYP2C19*2 G681A, CYP2C19*3 G636A, 2549delA, and CYP2D6*4 G1846A. For genotyping the CYP2D6*3 CYP2C9*1,*2,*3 and CYP2C19*1,*2,*3 SNPs, real-time PCR Fluorescence Resonance Energy Transfer (FRET) assays were performed using the CYP2C9 and CYP2C19 Mutation Detection Kits (Roche Diagnostics, Mannheim, Germany). The CYP2D6*1,*3 and *4 SNPs were genotyped using FRET assays (TIB MOLBIOL, Berlin, Germany) as described by Stamer et al.²⁴ and Müller et al.²⁵ on the LightCycler[®] (Roche Diagnostics, Mannheim, Germany)

Statistical analysis

Statistical analyses were performed with SPSS 15.0 (SPSS. Inc., Chicago, IL, USA) for Windows. In order to evaluate the association between the presence of pulmonary disease and the presence of CYP polymorphisms, odds ratios (ORs) with corresponding 95% confidence intervals (CI) were calculated. Actual allele distributions were compared against the expected frequencies calculated, using the Hardy-Weinberg equilibrium. Deviations from Hardy-Weinberg equilibrium were analysed using the chi-square test. A p-value of <0.05 (two sided) was considered to indicate statistical significance. A Bonferroni correction, to adjust for multiple comparisons, was applied were it was appropriate (p<0.01, indicating statistical significance).

Results

A summary of the characteristics of the DI-ILD, IPF and healthy volunteer population is shown in Table 4.1.

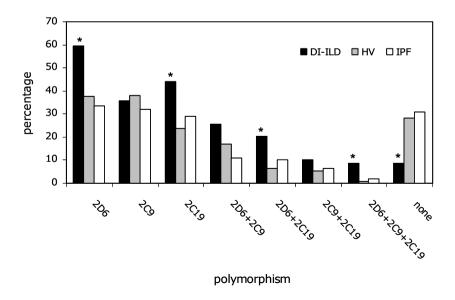
The subjects in the healthy volunteers group were younger than the subjects in the other two populations and used no medication. The age difference had no influence on the distribution of genetically variant genes, however, it does indicate that (multiple) drug use increases with age. The IPF control patients did use (multiple) drugs, but to a lesser extent than the DI-ILD patients. Table 4.1 also shows that of the patients in the ILD group, substantial larger percentages were receiving drugs that are metabolized by a polymorphic CYP enzyme system. For example, of the 43 patients who received a drug that should be metabolized by CYP2D6, 73% had a genetic polymorphism that would be likely to influence their ability to metabolize drugs efficiently.

Table 4.1 Baseline characteristics of the drug-induced interstitial lung disease (DI-ILD), idiopathic pulmonary fibrosis (IPF) and healthy volunteers (HV) populations.

Characteristic	DI-ILD	IPF	HV
Subjects (no.)	59	110	173
Male/female (no.)	28/31	62/48	78/95
Age (mean/range (y))	65.4/21-87	63.3/27-89	38.5/19-59
Percentages of above population	ons taking one or more dr	ugs metabolized by C	CYP polymorphic enzymes:
CYP2D6	73	18	0
CYP2C9	52	19	0
CYP2C19	88	17	0

CYP=cytochrome P450.

The percentages of individuals having one of the studied individual polymorphisms (CYP2D6, CYP2C9 and CYP2C19) or combinations in DI-ILD, healthy volunteer and IPF control patient groups, respectively, are shown in Figure 4.1.



Percentage of individual and combinations of cytochrome P450 (CYP) polymorphic genes Figure 4.1 in individuals with drug-induced interstitial lung disease (DI-ILD, n=59), idiopathic pulmonary fibrosis (IPF, n=110) and healthy volunteer (HV, n=173) groups. * p<0.05.

The genotype distribution for the three groups in this study showed that all results were generally consistent with the Hardy-Weinberg equilibrium (Table 4.2, upper panel, expected values not shown). ²⁶⁻²⁹ One exception was the CYP2D6 polymorphism of the healthy volunteer group, in which a small but significant deviation from the Hardy-Weinberg equilibrium was observed (p<0.01). Fewer heterozygotes (45 actually present in the healthy volunteer group versus 56 expected, calculated using the Hardy-Weinberg equilibrium) and more homozygotes (13 versus 7) were found. However, the allele frequencies in both control groups (healthy volunteers and IPF patients) were comparable with those in the reference populations from the literature (Table 4.2. lower panel). 26,27,29

Table 4.3 presents the results from the analyses that compared the proportions of persons with one ore more variant genes that express the polymorphic CYP enzymes between DI-ILD patients and both control groups. The proportion of persons without any allelic variants was only 8.5% among DI-ILD cases versus 30.9% and 29.5% in IPF patients and healthy volunteers. Furthermore, it appeared that there was a higher prevalence of combinations of variant genes within the DI-ILD patients compared with the IPF patients and healthy volunteers (Table 4.3, upper panel). Subjects without any CYP variant genes within the DI-ILD group and healthy volunteer group were used as reference group for the calculation of ORs associated with the presence of one or more of the studied genetic polymorphisms. Strong and significant associations were found comparing the prevalence of CYP2D6 and CYP2C19 genetic variants in DI-ILD patients with those of the healthy volunteers. The ORs calculated were 5.80 (95% CI: 2.17-15.4, p<0.001) for CYP2D6, 3.25 (95% CI: 1.18-8.86, p=0.026) for CYP2C9 and 6.47 (95% CI: 2.35-17.7, p<0.001) for CYP2C19, respectively (Table 4.3). For the combined genotypes with variant CYP alleles, significant ORs were found of 4.91, 14.0, 6.80 and 40.8 (Table 4.3).

Similar results were found comparing the prevalence of variant CYP genes in DI-ILD patients with the IPF control patients. These analyses, as shown in Table 4.3, also resulted in significant associations between the development of DI-ILD and the presence of CYP genetic variants.

The percentage of DI-ILD patients having a CYP variant gene and receiving one or more drug(s) metabolized by a polymorphic CYP enzyme was 87% (47/54), compared with only 16% (12/76) of the IPF patients.

Table 4.4 lists the (suspected) causative drugs, the CYP enzymes involved in the metabolism of the mentioned drugs, the number of DI-ILD patients involved (with and without variant CYP genes) and also shows which of the variant forms of CYP genes were found. The last column lists the number of literature references mentioned on www.pneumotox.com, concerning the stated causative drug in relation to lung disease.

Table 4.2 Genotype distribution and allele frequencies of cytochrome P450 (CYP) isoforms.

- Population ^a	<i>2D6</i>				2C9				2C19			
	DI-ILD IPF		Η	U	DI-ILD	IPF	Α	U	DI-ILD	IPF	¥	U
No. (%)	29	110	173	765	59	110	173	121	59	110	173	736
Wild type ^b	26 (44.1)	73 (66.4)	115 (66.4)	26 (44.1) 73 (66.4) 115 (66.4) 490 (64.2) 38 (64.4) 75 (68.2) 107 (61.8) 82 (67.8) 33 (55.9) 78 (70.9) 132 (76.3) 554 (75.3)	38 (64.4)	75 (68.2)	107 (61.8)	82 (67.8)	33 (55.9)	78 (70.9)	132 (76.3)	554 (75.3)
Heterozygous ^c 26 (44.1) 30 (27.3) 45 (26.0) 233 (30.3) 17 (28.8) 33 (30.0) 62 (35.8) 36 (29.8) 22 (37.3) 26 (23.6) 35 (20.2) 163 (22.1)	26 (44.1)	30 (27.3)	45 (26.0)	233 (30.3)	17 (28.8)	33 (30.0)	62 (35.8)	36 (29.8)	22 (37.3)	26 (23.6)	35 (20.2)	163 (22.1)
Homozygous ^d 7 (11.9) 7	7 (11.9)	7 (6.4)	13 (7.5)	(6.4) 13 (7.5) 42 (5.5) 4 (6.8) 2 (1.8) 4 (2.3)	4 (6.8)	2 (1.8)	4 (2.3)	3 (2.5)	4 (6.8)	6 (5.5)	6 (3.5) 19 (2.6)	19 (2.6)
Allele (%)												
*1	66.1	0.08	79.5	79.3	78.8	83.2	79.8	82.2	74.6	82.7	86.4	86.5
*2	NA	ΝΑ	N A	Ą	12.7	8.2	13.8	10.0	25.4	17.3	13.6	13.3
*3	5.1	1.4	1.4	1.8	8.5	9.8	6.4	7.4	0	0	0	0.2
*4	28.8	18.6	19.1	18.4	NA	NA	NA	NA	NA	NA	NA	NA

^a CYP2D6, CYP2C9 and CYP2C19 genetic polymorphisms in DI-ILD patients (n=59); IPF control patients (n=110), healthy volunteers (n=173) and C populations as a reference. For CYP2D6 and CYP2C19, allele frequencies from Tamminga et al. 26 and Sachse et al. 29 were used and for CYP2C9, allele frequencies from Allabi et al.²⁷ were used.

 $^{^{}m b}$ Wild type (fully functional enzyme).

Heterozygote variant (partially functional enzyme).
 Homozygote variant (severely compromised or non-functional enzyme).

C=Caucasian populations from literature; DI-ILD=drug-induced interstitial lung disease; HV=healthy volunteers; IPF=idiopathic pulmonary fibrosis; NA=not applicable.

Comparison of the percentages of subjects with one or more variant cytochrome P450 (CYP) genes between drug-induced interstitial lung disease (DI-ILD) patients (n=59), healthy volunteers (HV; n=173) and idiopathic pulmonary fibrosis (IPF) control patients (n=110), using individuals without any CYP variants as a reference group. Table 4.3

CYP gene	2D6	2C9	2C19	2D6+2C9	2D6+2C19	2C9+2C19	2D6+2C9+2C19	None
DI-ILD n (%)	33 (55.9)	21 (35.6)	26 (44.1)	13 (22.0)	11 (18.6)	6 (10.2)	4 (6.8)	5 (8.5)
IPF n (%)	37 (33.6)	35 (31.8)	32 (29.1)	12 (10.9)	11 (10.0)	7 (6.4)	2 (1.8)	34 (30.9)
HV n (%)	58 (33.5)	66 (38.2)	41 (23.7)	27 (15.6)	8 (4.6)	9 (5.2)	1 (0.6)	51 (29.5)
DI-ILD vs HV								
Odds ratio	5.80	3.25	6.47	4.91	14.0	6.80	40.8	1.00
95% CI	2.17-15.4	1.18-8.86	2.35-17.7	1.63-14.6	3.96-46.6	1.79-26.0	4.82-318.4	AN
p-value	<0.001*	0.026	<0.001*	0.007*	<0.001*	0.008*	0.001*	NA
DI-ILD vs IPF								
Odds ratio	6.07	4.08	5.53	7.37	6.80	5.83	13.6	1.00
95% CI	2.18-16.7	1.42-11.6	1.94-15.6	2.23-24.2	1.99-23.1	1.45-23.6	2.22-82.1	NA
p-value	<0.001*	0.010	0.001*	0.001*	0.002*	0.019	0.010	NA

NA=not applicable; vs=versus; *=significantly different (p<0.01) - a Bonferroni correction was applied.

Table 4.4 List of potentially causative drugs and the cytochrome P450 (CYP) genes and variant genotypes expressing enzymes involved in their metabolism.

Causative drug	CYP gene ^a	Allelic variants present	Drug+	No variant gene+	Literature ^d
			variant gene ^b	inhibitor ^c	
Acenocoumarol	2C9	*1/*2 *2/*3	5	7	4
	2C19	*1/*2 *2/*2	8	2	4
Acetaminophen	2D6	*1/*4	2	2	11
	2C9	*1/*2 *2/*3	2	1	11
Amiodarone	2D6	*1/*4 *1/*3 *3/*4 *4/*4	8	7	276
	2C19	*1/*2	6	9	276
Amitriptyline	2D6	*1/*4	2		4
	2C19	*1/*2	1		4
Captopril	2D6	*3/*4	1	1	27
Carbamazepine	2C9			1	53
Carvedilol	2D6	*1/*3	1	1	2
Cotrimoxazol	2C9			1	11
Cyclophosphamid		*1/*2	1		65
Diazepam	2C19	*1/*2	1		_
Diclofenac	2C9	*3/*3	1	1	3
	2C19	*1/*2	1		3
Fenfluramine	2D6	*1/*3	1		39
Flecainide	2D6	*1/*3	1		10
Fluoxetine	2D6	*1/*4 *1/*3	2	1	8
	2C19	*1/*2	2		8
Haloperidol	2D6			1	2
Ibuprofen	2C9	*1/*2	1	1	6
	2C19	*1/*2	1		6
Methotrexate				2	117
Metoprolol	2D6	*1/*4 *4/*4 *1/*3	10	1	2
Montelukast	2C9	*1/*2	1		6
Morphine	2D6	*1/*4	1		16
Naproxen	2C9	*1/*2	1	1	14
Omeprazol	2C19	*1/*2	4	5	1
Pantoprazol	2C19	*1/*2 *2/*2	5	2	
Paroxetine	2D6	*1/*4	2	2	2
Propranolol	2D6	*1/*4 *3/*4	1	1	11
Ranitidine	2D6	*1/*4	2	1	
	2C19	*1/*2	2	1	
Risperidone	2D6	*1/*4	1		1
Tamsulosin	2D6	*1/*3 *3/*4	3	1	
Temazepam	2C19	*1/*2 *2/*2	4		
Tramadol	2D6	*1/*4	1		2
Valsartan	2C9	*1/*3 *1/*2 *2/*3	3	1	3
Venlafaxine	2D6	*1/*4	2		5
	2C19	*1/*2	1		5

^a Gene expressing CYP enzyme involved in the metabolism of the drug.

^b Total number of patients with a variant *CYP* gene receiving the drug.

 $^{^{\}mathrm{c}}$ Total number of patients receiving the drug, without a variant CYP gene, but with the drug being associated with an inhibition of the CYP enzyme involved in the metabolism of the drug.

^d Total number of articles at www.pneumotox.com referring to the mentioned drug.

Discussion

To the best of our knowledge, this is the first study indicating that DI-ILD may be attributable to a reduced metabolic capacity by CYP enzymes. According to our results, there seems to be an association between having one or more CYP genetic variants that may lead to reduced metabolism, receiving drugs that are metabolized inadequately by the affected system and the development of DI-ILD. Looking at the ORs, ranging from 3.25 to 40.8, patients with variant CYP enzymes appear to be at a substantially greater risk of developing a DI-ILD when prescribed multiple drugs. These findings strengthened our presumption that inadequate drug metabolism predispose an individual for the development of DI-ILD. In 91.5% of the studied DI-ILD patients at least one CYP variant gene was present, whereas this percentage was about 70% in the control populations. In the DI-ILD group the prevalence of having more than one CYP variant gene was also substantially higher. The results support the potential usefulness of CYP genotyping in selecting appropriate drugs or dosages of drugs and avoiding subsequent serious adverse effects.

CYP isoenzymes have been detected in animal as well as in human lung tissues. 30 It is generally agreed that the CYP super family of enzymes forms the first step in the inactivation and elimination of numerous drugs by oxidation and reduction. Considering the fact that bio-activation by CYP enzymes plays an important role in human drug toxicity, polymorphisms in the CYP450 enzyme system may result in large inter-individual variations in the metabolism and toxicity of xenobiotics.

Given the ever-increasing number of patients, especially seriously ill and/or elderly, who take more than one drug, also often metabolized by the same CYP enzyme, the inherent problems of drug-induced toxicity are alarming.31 Physicians should therefore be alert to the possibility that a drug-induced pulmonary reaction may originate from an inappropriate metabolism, especially in case of multiple drug use. The results in this study corroborate that genotyping a patient before drug initiation could lead to a more tailor-made dosing schedule that might protect the patient from the development of serious side effects at the start of the pharmacotherapy. Trial-and-error approaches could be reduced this way.³²

In the present study we focused on CYP polymorphisms. The total 'genetic profile' of an individual patient should include genes expressing further polymorphic enzymes and other proteins involved in drug metabolism and response. For example, in the case of azathioprine indication, also used as treatment for IPF, testing for thiopurine methyltransferase (TPMT) variants

azathioprine metabolism is advised before involved in the treatment.³³⁻³⁵ In the US, drug labels for azathioprine now include information on TPMT polymorphisms and recommend determining patients' phenotype or genotype prior to drug treatment.³⁶

The finding that 87% of the studied DI-ILD patients received one or more drug(s) metabolized by an affected CYP metabolism route is consistent with the assumption that the interstitial lung reactions were drug-induced and that the case group in this study was well defined. In the other cases drug-drug interactions may have been responsible for the toxic drug effects or other pharmacogenetic factors might be involved, such as reduced TPMT activity involved in methotrexate metabolism. 37,38

There are an increasing number of examples where pharmacogenetic studies have indicated that a genetic test prior to treatment may be useful either for setting the individual dose or making a decision to use a particular drug.³⁹ The ability to identify individuals who are susceptible to ADRs has the potential to reduce the personal and population costs of drug-related morbidity.8

In this retrospective study, the healthy volunteers used no medication and among the IPF group patients used fewer drugs compared with the DI-ILD patients. However, this does not imply that the healthy control patients and IPF patients are not at risk of developing a drug-induced pulmonary reaction. Persons with more than one CYP polymorphism and/or other relevant polymorphisms may be susceptible to develop DI-ILD when (multiple) drugs are prescribed. 9,35,37 To answer the question whether persons with one or more of these polymorphisms will develop a DI-ILD, whenever future drug prescription is mandatory, needs follow-up. However, since the CYP polymorphisms of these patients are already known, this can be taken into consideration when prescribing, thus avoiding (possible) adverse drug reactions. Moreover, a prospective case-control study deriving both treated patients and control patients from one population source of multi-drug users would be useful, in order to evaluate the cost effectiveness of the introduction of pharmacogenetic testing into routine healthcare. Such studies will help to identify factors that increase the risk of unwanted outcomes from drug therapy. They will also help to establish in which circumstances genotyping should be performed prior to commencing drug treatment and in tailoring drug treatment for individual patients.^{8,9}

Conclusion

This study indicates that the presence of CYP variant genotypes appeared to be a substantial susceptibility risk factor in the development of drug-induced pulmonary adverse events. Therefore, genotyping prior to drug prescription may be clinical useful for the prediction and prevention of drug-induced pulmonary toxicity, especially in case of multiple drug use, where prior genotyping or phenotyping has the potential to contribute to the patients' safety. Both clinical and genetic risk stratification (pharmacogenomics) may lead to more accurate prevention of drug-induced damage in the future.

References

- McLeod HL, Evans WE. Pharmacogenomics: unlocking the human genome for better drug therapy. Annu Rev Pharmacol Toxicol. 2001; 41:101-21.
- 2. Camus P, Fanton A, Bonniaud P, Camus C, Foucher P. Interstitial lung disease induced by drugs and radiation. Respiration. 2004; 71:301-26.
- Weinshilboum R. Inheritance and drug response. N Engl J Med. 2003; 348:529-37. 3.
- Yang IV, Burch LH, Steele MP, Savov JD, Hollingsworth JW, McElvania-Tekippe E, 4. Berman KG, Speer MC, Sporn TA, Brown KK, Schwarz MI, Schwartz DA. Gene expression profiling of familial and sporadic interstitial pneumonia. Am J Respir Crit Care Med. 2007: 175:45-54.
- 5. Selman M, Pardo A, Barrera L, Estrada A, Watson SR, Wilson K, Aziz N, Kaminski N, Zlotnik A. Gene expression profiles distinguish idiopathic pulmonary fibrosis from hypersensitivity pneumonitis. Am J Respir Crit Care Med. 2006; 173:188-98.
- 6. Wilkinson GR. Drug metabolism and variability among patients in drug response. N Engl J Med. 2005; 352:2211-21.
- Nebert DW, Russell DW. Clinical importance of the cytochromes P450. Lancet. 7. 2002: 360:1155-62.
- Clark DW, Donnelly E, Coulter DM, Roberts RL, Kennedy MA. Linking pharmacovigilance with pharmacogenetics. Drug Saf. 2004; 27:1171-84.
- Jaquenoud Sirot E, van der Velden JW, Rentsch K, Eap CB, Baumann P. Therapeutic 9. drug monitoring and pharmacogenetic tests as tools in pharmacovigilance. Drug Saf. 2006; 29:735-68.
- 10. Nemery B, Bast A, Behr J, Borm PJ, Bourke SJ, Camus PH, De Vuyst P, Jansen HM, Kinnula VL, Lison D, Pelkonen O, Saltini C. Interstitial lung disease induced by exogenous agents: factors governing susceptibility. Eur Respir J Suppl. 2001; 32:30s-42s.
- 11. Wormhoudt LW, Commandeur JN, Vermeulen NP. Genetic polymorphisms of human N-acetyltransferase, cytochrome P450, glutathione-S-transferase, and epoxide hydrolase enzymes: relevance to xenobiotic metabolism and toxicity. Crit Rev Toxicol. 1999; 29:59-124.
- 12. Higenbottam T, Kuwano K, Nemery B, Fujita Y. Understanding the mechanisms of drug-associated interstitial lung disease. Br J Cancer. 2004; 91 Suppl 2:S31-7.
- 13. Hukkanen J, Pelkonen O, Raunio H. Expression of xenobiotic-metabolizing enzymes in human pulmonary tissue: possible role in susceptibility for ILD. Eur Respir J Suppl. 2001; 32:122s-6s.
- 14. Castell JV, Donato MT, Gomez-Lechon MJ. Metabolism and bioactivation of toxicants in the lung. The in vitro cellular approach. Exp Toxicol Pathol. 2005; 57 Suppl 1:189-204.
- 15. Bast A. Is formation of reactive oxygen by cytochrome P-450 perilous and predictable? Trends Pharmacol Sci. 1986; 7:266-70.
- 16. Bast A, Haenen GR, Doelman CJ. Oxidants and antioxidants: state of the art. Am J Med. 1991; 91:2S-13S.
- 17. Guidice JM, Marez D, Sabbagh N, Legrand-Andreoletti M, Spire C, Alcaïde E, Lafitte JJ, Broly F. Evidence for CYP2D6 expression in human lung. Biochem Biophys Res Commun. 1997; 241:79-85.
- 18. Wilschut FA, Cobben NA, Thunnissen FB, Lamers RJ, Wouters EF, Drent M. Recurrent respiratory distress associated with carbamazepine overdose. Eur Respir J. 1997; 10:2163-5.
- 19. Drent M, Singh S, Gorgels AP, Hansell DM, Bekers O, Nicholson AG, van Suylen RJ, du Bois RM. Drug-induced pneumonitis and heart failure simultaneously associated with venlafaxine. Am J Respir Crit Care Med. 2003; 167:958-61.

- 20. Drent M, Jacobs JA, Cobben NA, Costabel U, Wouters EF, Mulder PG. Computer program supporting the diagnostic accuracy of cellular BALF analysis: a new release. *Respir Med.* 2001; 95:781-6.
- 21. Lindell RM, Hartman TE. Chest imaging in iatrogenic respiratory disease. *Clin Chest Med.* 2004; 25:15-24.
- 22. Wijnen PA, Op den Buijsch RA, Cheung SC, van der Heijden J, Hoogtanders K, Stolk LM, van Dieijen-Visser MP, Neef C, Drent M, Bekers O. Genotyping with a dried blood spot method: A useful technique for application in pharmacogenetics. *Clin Chim Acta*. 2008;388:189-91.
- 23. American Thoracic Society. Idiopathic pulmonary fibrosis: diagnosis and treatment. International consensus statement. American Thoracic Society (ATS), and the European Respiratory Society (ERS). *Am J Respir Crit Care Med.* 2000; 161:646-64.
- 24. Stamer UM, Bayerer B, Wolf S, Hoeft A, Stüber F. Rapid and reliable method for cytochrome P450 2D6 genotyping. *Clin Chem.* 2002; 48:1412-7.
- 25. Muller B, Zopf K, Bachofer J, Steimer W. Optimized strategy for rapid cytochrome P450 2D6 genotyping by real-time long PCR. *Clin Chem.* 2003; 49:1624-31.
- 26. Tamminga WJ, Wemer J, Oosterhuis B, de Zeeuw RA, de Leij LF, Jonkman JH. The prevalence of CYP2D6 and CYP2C19 genotypes in a population of healthy Dutch volunteers. *Eur J Clin Pharmacol.* 2001; 57:717-22.
- Allabi AC, Gala JL, Desager JP, Heusterspreute M, Horsmans Y. Genetic polymorphisms of CYP2C9 and CYP2C19 in the Beninese and Belgian populations. Br J Clin Pharmacol. 2003; 56:653-7.
- 28. Bravo-Villalta HV, Yamamoto K, Nakamura K, Bayá A, Okada Y, Horiuchi R. Genetic polymorphism of CYP2C9 and CYP2C19 in a Bolivian population: an investigative and comparative study. *Eur J Clin Pharmacol.* 2005; 61:179-84.
- 29. Sachse C, Brockmoller J, Bauer S, Roots I. Cytochrome P450 2D6 variants in a Caucasian population: allele frequencies and phenotypic consequences. *Am J Hum Genet.* 1997; 60:284-95.
- 30. Zhang JY, Wang Y, Prakash C. Xenobiotic-metabolizing enzymes in human lung. *Curr Drug Metab.* 2006; 7:939-48.
- 31. Ensom MH, Chang TK, Patel P. Pharmacogenetics: the therapeutic drug monitoring of the future? *Clin Pharmacokinet*. 2001; 40:783-802.
- 32. Reitman ML, Schadt EE. Pharmacogenetics of metformin response: a step in the path toward personalized medicine. *J Clin Invest.* 2007; 117:1226-9.
- 33. Schutz E, von Ahsen N, Oellerich M. Genotyping of eight thiopurine methyltransferase mutations: three-color multiplexing, "two-color/shared" anchor, and fluorescence-quenching hybridization probe assays based on thermodynamic nearest-neighbor probe design. *Clin Chem.* 2000; 46:1728-37.
- 34. Baker DE. Pharmacogenomics of azathioprine and 6-mercaptopurine in gastroenterologic therapy. *Rev Gastroenterol Disord.* 2003; 3:150-7.
- 35. Bakker JA, Bierau J, Drent M. Therapeutic regimens in interstitial lung disease guided by genetic screening: fact or fiction? *Eur Respir J.* 2007; 30:821-2.
- 36. Daly AK. Individualized drug therapy. Curr Opin Drug Discov Devel. 2007; 10:29-36.
- 37. Campalani E, Arenas M, Marinaki AM, Lewis CM, Barker JN, Smith CH. Polymorphisms in folate, pyrimidine, and purine metabolism are associated with efficacy and toxicity of methotrexate in psoriasis. *J Invest Dermatol.* 2007; 127:1860-7.
- Wessels JA, de Vries-Bouwstra JK, Heijmans BT, Slagboom PE, Goekoop-Ruiterman YP, Allaart CF, Kerstens PJ, van Zeben D, Breedveld FC, Dijkmans BA, Huizinga TW, Guchelaar HJ. Efficacy and toxicity of methotrexate in early rheumatoid arthritis are associated with single-nucleotide polymorphisms in genes coding for folate pathway enzymes. Arthritis Rheum. 2006; 54:1087-95.
- 39. Shurin SB, Nabel EG. Pharmacogenomics--ready for prime time? *N Engl J Med.* 2008; 358:1061-3.